



Aroma Profile of Essential Oils of *Solenostemon monostachyus* P. Beauv from Nigeria

Emmanuel E. Essien^{1,3*}, Paul S. Thomas², Mohammad I. Choudhary³

¹Department of Chemistry, University of Uyo, Uyo 520101, Nigeria

²Department of Pharmacognosy and Natural Medicine, Faculty of Pharmacy, University of Uyo, Uyo 520101, Nigeria

³Hussain Ebrahim Jamal Research Institute of Chemistry, International Centre for Chemical and Biological Sciences, University of Karachi, Karachi 75270, Pakistan

Article Information

Received 5 January 2017

Received in revised form 24 April 2017

Accepted 25 April 2017

Keywords:

Solenostemon monostachyus,
Lamiaceae,
essential oil,
gas chromatography-mass
spectrometry

Corresponding Author:

E-mail : emmaflowus1@yahoo.co.uk

Mob.: +234-8033683424

Abstract

This study was conducted to evaluate the volatile oils constituents of *Solenostemon monostachyus* P. Beauv, an aromatic non-conventional vegetable. The essential oils isolated by hydrodistillation from leaves, stem, floral and aerial parts of *S. monostachyus* (P.Beauv.) Briq. were investigated by gas chromatography-mass spectrometry (GC-MS). A total of fifteen constituents were identified and characterized by the high amount of sesquiterpenoid (43.13-91.21%). The leaf oil comprised mainly of β -caryophyllene (71.42%), α -caryophyllene (7.0%), 1-octen-3-ol (6.96%) and caryophyllene oxide (6.67%); the stem oil consisted of β -caryophyllene (46.75%), α -caryophyllene (20.22%), caryophyllene oxide (10.45%) and 1-octen-3-ol (10.31%); the predominant compounds in the floral oil were 1-octen-3-ol (40.24%), β -caryophyllene (18.94%) and α -caryophyllene (16.98%); while β -caryophyllene (27.43%), caryophyllene oxide (24.83%) and α -caryophyllene (12.9%) were the abundant components of the aerial oil. The chemical composition of *S. monostachyus* essential oils from Nigeria is reported for the first time.

1 Introduction

Volatile oils (also known as essential oils) are concentrated hydrophobic liquid consisting of volatile aroma compounds from plants. Volatile oils utility range from aromatherapy, household cleaning products, personal beauty care, and natural medicine treatments¹. *Solenostemon monostachyus* P. Beauv (family Lamiaceae) is an essential oil bearing plant, and an important edible herb that is widespread in West and Central Africa. The plant is an erect, branched annual weed with a long inflorescence of violet flowers (Fig. 1). It is slightly succulent, aromatic and grows up to 100 cm tall². The leaves are used to treat dysmenorrhoea, haematuria, female sterility, rheumatism, foot infections, convulsions, fever, hypertension, stomach ulcer, hemorrhoid and snakebites; the plant has many ritual uses, especially related to pregnancy³⁻⁶. Phytochemical studies on *S. monostachyus* leaves afforded the isolation of diterpenoids⁷ and essential oil². Research has also shown that the leaf extracts of *S. monostachyus* exhibits antioxidant^{8,9}, antihypertensive¹⁰,

antimicrobial¹¹ and antiulcerogenic activities¹². There is paucity of data on the essential oil composition of *S. monostachyus*. Published data on *S. monostachyus* essential oil is limited to the Cameroon leaf oil sample².



Fig. 1: *Solenostemon monostachyus* plant

Therefore, in continuation of a systematic analysis of essential oil constituents of relatively poorly studied aromatic medicinal plants¹³, we report for the first time, the composition of *S. monostachyus* essential oils from Nigeria.

2 Materials and methods

2.1. Plant Material

The mature *S. monostachyus* plants were collected from the wild in Uyo Local Government Area of Akwa Ibom State, Nigeria, in the month of May 2015. Plant samples were identified and authenticated by a taxonomist, M. E. Bassey, Department of Botany and Ecological Studies, University of Uyo, where voucher specimens were deposited. The essential oils were obtained by hydrodistillation (4 h) of the fresh plant parts using a Clevenger-type apparatus in accordance with the British Pharmacopoeia¹⁴. The oils were dried over sodium sulfate and kept in refrigeration (4 °C) after estimation of percentage yield.

2.2 Gas Chromatography - Mass Spectrometry (GC – MS)

The essential oils were subjected to GC-MS analysis on an Agilent system consisting of a model 7890N gas chromatograph, a model mass detector Triple Quad 7000A in EI mode at 70 eV (*m/z* range 40–600 amu) (Agilent Technologies, Santa Clara, CA, USA), and an Agilent ChemStation data system. The GC column was an HP-5ms fused silica capillary with a (5% phenyl)-methyl polysiloxane stationary phase (30 m x 250 µm x 0.25 µm). The carrier gas was helium with a column head pressure of 9.7853 psi and flow rate of 1.2 mL/min. Inlet temperature and MSD detector temperature was 250 °C.

The GC oven temperature program was used as follows: 50 °C initial temperature, held for 5 min; increased at 6 °C/min to 190 °C for 20 mins; increased 7 °C/min to 290 °C for 15 mins; increased 7 °C/min to 300 °C for 10 mins. The sample was dissolved in CH₂Cl₂, and 2 µL was injected (split ratio 10:1; split flow 12 mL/min).

The components were identified by comparison of their mass spectra with NIST 1998 library data of the GC-MS system as well as by comparison of their retention indices (RI) with the relevant literature data¹⁵. The relative amount of each individual component of the essential oil was expressed as the percentage of the peak area relative to the total peak area. RI value of each component was determined relative to the retention times of a homologous n-alkane series with linear interpolation on the HP-5ms column.

3 Result and Discussion

The yields of the leaf, stem, floral and aerial essential oils of *S. monostachyus* were 0.2%, 0.13%, 0.16% and 0.2%, respectively. The aroma profile of *S. monostachyus* volatile oils are presented in Table 1. The essential oil composition of the different plant parts was distinct, quantitatively and qualitatively.

A total of fifteen (15) constituents were identified accounting for 89.72% to 98.17% of the oils content and characterized by the high amount of sesquiterpenoid (43.13-91.21%). Monotepenoid was not detected in the leaf and stem oils, however, relatively low in the floral and aerial oils (4.29 % and 9.81% respectively). The leaf oil comprised mainly of β-caryophyllene (71.42%), α-caryophyllene (7.0%), 1-octen-3-ol (6.96%) and caryophyllene oxide (6.67%); the stem oil consisted of β-caryophyllene (46.75%), α-caryophyllene (20.22%), caryophyllene oxide (10.45%) and 1-octen-3-ol (10.31%); the predominant compounds in the floral oil were 1-octen-3-ol (40.24%), β-caryophyllene (18.94%) and α-caryophyllene (16.98%); while β-caryophyllene (27.43%), caryophyllene oxide (24.83%) and α-caryophyllene (12.9%) were the abundant components of the aerial oil. α-caryophyllene, β-caryophyllene and caryophyllene oxide were common constituents of the studied essential oils (Fig. 2).

Analysis also revealed the high amount of 1-octen-3-ol in the floral oil sample (40.24%) relative to other studied plant parts. The leaf essential oil of *S. monostachyus* from Cameroon is reported to contain β-pinene (13.0%), oct-1-en-3-ol (12.6%), β-caryophyllene (6.9%), octan-3-ol (6.8%) and (*E,E*)-α-farnesene (6.2%) as major constituents².

β-Caryophyllene occurred in a very high amount in the Nigerian leaf sample compared with the Cameroon sample while β-pinene was not detected in the Nigerian oil. However, several constituents such as 1-octen-3-ol, β-caryophyllene, among others, are common to both leaf oils of different geographical origins. It is worthy of note that caryophyllene rich essential oils such as essential oil of *Stachys cretica* (β-caryophyllene, 51.0%) and β-caryophyllene are reported to exhibit strong antimicrobial activity, particularly against *P. aeruginosa* and *B. subtilis*¹⁶ and cytotoxic activity¹⁷.

4 Conclusions

The volatile constituents of *S. monostachyus* have been analyzed and identified. The leaf, stem, floral and aerial essential oils of *S. monostachyus* from Nigeria displayed significant quantitative and qualitative chemical profiles characterized mainly of sesquiterpene hydrocarbons.

5 Acknowledgements

The chemical analysis of the essential oils was made possible by funding through the ICCBS-TWAS Post-doctoral Fellowship awarded to Dr. Emmanuel Essien and utilized at the H.E.J Research Institute of Chemistry, International Centre for Chemical and Biological Sciences (ICCBS), University of Karachi, Karachi, Pakistan.

6 Conflicts of Interest

The authors declare no conflict of interest.

7 Author Contributions

UK J Pharm & Biosci, 2017: 5(2); 35

EEE and PST conceived and designed the experiments; EEE and PST performed the experiments; EEE wrote the manuscript; MIC supervised the experiments.

Table 1: Volatile constituents of *Solenostemon monostachyus*

RI ^a	Compound ^{b,c}	Leaf	Stem	Flower	Aerial	QI ^d
980	1-Octen-3-ol	6.96	10.31	40.24	-	95
982	β -Pinene	-	-	-	6.96	95
1013	δ -3-Carene	-	-	1.27	1.23	92
1067	(E)-2-Octen-1-ol	-	-	-	5.96	96
1103	β -Linalool	-	-	3.02	-	96
1200	Dodecane	-	-	2.45	-	93
1327	Myrtenyl acetate	-	-	-	1.62	97
1377	α -Copaene	-	-	2.17	2.26	94
1419	β -Caryophyllene	71.42	46.75	18.94	27.43	95
1482	Germacrene D	-	-	-	2.21	97
1524	δ -Cadinene	3.31	-	1.15	2.08	98
1564	α -Caryophyllene	7.0	20.22	16.98	12.90	95
1582	Caryophyllene oxide	6.67	10.45	3.89	24.83	97
1585	Globulol	2.81	-	-	-	94
1645	δ -Cadinol	-	-	-	2.24	96
	Monoterpene hydrocarbons	-	-	1.27	8.19	
	Oxygenated monoterpene	-	-	3.02	1.62	
	Monoterpenoid	-	-	4.29	9.81	
	Sesquiterpene hydrocarbons	81.73	66.97	39.24	46.88	
	Oxygenated sesquiterpene	9.48	10.45	3.89	27.07	
	Sesquiterpenoid	91.21	77.42	43.13	73.95	
	Aliphatic alcohol	6.96	10.31	40.24	5.96	
	Aliphatic hydrocarbons	-	-	2.45	-	
	Total	98.17	87.73	90.11	89.72	

^aRI, calculated retention indices; ^bOrder of elution on HP-5ms capillary column; ^cIdentification by comparison of the mass spectral and retention index data;

^dQI, 'quality index', reflects the fit comparison of experimental mass spectrum and NIST library mass spectrum; - = not detected.

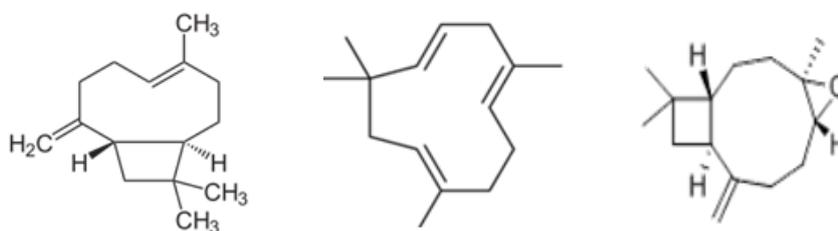


Fig. 2: Common constituents of *S. monostachyus* essential oils: β -caryophyllene, α -caryophyllene and caryophyllene oxide.

8 References

- Ryman D. (1984). *The Aromatherapy Handbook: The Secret Healing Power of Essential Oils*. Chapter 3, Century Publishing Co. Ltd., 1984.
- Mvé-Mba CE, Menut C, Lamaty G, Zollo, PHA, Tchoumboungang F, Bessière JM. Aromatic plants of tropical central Africa. Part XIX. Volatile components from leaves of two Lamiaceae from Cameroon: *Leucas deflexa* Hook and *Solenostemon monostachyus* (P. Beauv.) Briq. *Flav. Fragr. J.* 1994; 9(6): 315-317.
- Lemmens RHMJ. *Solenostemon monostachyus* (P. Beauv.) Briq. In: Grubben GJH & Denton OA (Eds). *PROTA 2: Vegetables/Légumes*. [CD-Rom]. PROTA, Wageningen, Netherlands, 2004.
- Ajibesin KK, Ekpo BA, Bala DN, Essien EE, Adesanya SA. Ethnobotanical survey of Akwa Ibom State of Nigeria. *J. Ethnopharmacol.* 2008; 115: 387– 408.
- Adebayo JO, Krettli AU. Potential antimalarials from Nigerian plants: A review. *J. Ethnopharmacol.* 2011; 133: 289–302.
- Koffi N, Marie –Solange T, Emma AA, Noel ZG. Ethnobotanical study of plants used to treat arterial hypertension in traditional medicine, by Abbey and Krobou population of Agboville (Cote d'ivoire). *Eur. J. Sci. Res.* 2009; 35: 85-98.
- Toshio M, Peter R, Conrad HE. Structures of six coleons (diterpenoids) from *Solenostemon monostachyus* (P. Beauv.) Briq. (Labiatae). *Helvetica Chimica Acta* 1980; 63: Fasc.1-Nr.9 10.
- Datte, JY, Kpahe F., Offoumou AM. Acute toxicity and antioxidant activity of hydroethanolic extract of *Solenostemon monostachyus* P. Beauv. Leaves. *J. Compl. Integr. Med.* 2010; 7: Art. 45.
- Okoko T, Ere D. Antioxidant activities of *Solenostemon monostachyus* leaf extract using *in vitro* methods. *Sci. Res. Essays* 2012; 7(6): 621-626.
- Fidele KZ, Andre KB, Yao DJ, Michel OA. Action of hydroethanolic leaves extract of *Solenostemon monostachyus* (Lamiaceae) on cardiovascular system of mammals: blood pressure lowering effects. *Int. J. Pharm. Biol. Sci.* 2012; 2(3): 310-320.
- Ekundayo EO, Ezeogu LI. Evaluation of antimicrobial activities of extracts of five plants used in traditional medicine in Nigeria. *Intern. J. Trop. Med.* 2006; 1: 93-96.
- Amazu LU, Antia BS, Okokon JE. Antiulcerogenic activity of *Solenostemon monostachyus*. *The J. Phytopharmacol.* 2015; 4(2): 97-101.
- Thomas PS, Essien EE, Ntuk SJ, Choudhary MI. *Eryngium foetidum* L. essential oils: chemical composition and antioxidant capacity. *Medicines* 2017; 4(24): 1-7, doi: 10.3390/medines4020024.
- Medicines and Healthcare products Regulatory Agency (MHRA). *British Pharmacopoeia*, Vol. II; H.M. Stationery Office, Pharmaceutical Press: London, UK, 1980.
- Adams RP. *Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry*, 4th ed.; Allured Publishing Corp.: Carol Stream, IL, USA, 2007.
- Oztürk M, Duru ME, Aydoğmus-Oztürk F, Harmandar M, Mahliçli M, Kolak U, Ulubelen A. GC-MS analysis and antimicrobial activity of essential oil of *Stachys cretica* subsp. *smyrnaea*. *Nat. Prod. Commun.* 2009; 4: 109–114.
- Jun NJ, Mosaddik A, Moon JY, Jang K, Lee D, Ahn KS *et al.* Cytotoxic activity of β -caryophyllene oxide isolated from jeju guava (*Psidium cattleianum* Sabine) Leaf. *Rec. Nat. Prod.* 2011; 5(3): 242-246.